


The Impact of *BCR-ABL1* Transcript Type on Tyrosine Kinase Inhibitor Responses and Outcomes in Patients With Chronic Myeloid Leukemia

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Although the majority of patients with chronic myeloid leukemia do well with treatment with tyrosine kinase inhibitors (TKIs), some patients still have inferior outcomes. There are many factors that might play a part, including the different *BCR-ABL1* transcript types at baseline. The current study was performed to determine the possible impact of different transcripts on the treatment responses and outcomes of patients with chronic myeloid leukemia who are receiving TKI therapy. The authors performed a systematic literature search by using the terms “b2a2/b3a2,” “e13a2/e14a2,” or “transcript type.” e14a2 was the more common transcript type. The majority of the studies demonstrated no significant difference regarding age, sex, leukocyte counts, and hemoglobin levels between patients with the e13a2 and e14a2 transcripts. However, in approximately one-half of the studies, the e14a2 transcript was associated with higher platelet counts. Almost no studies demonstrated a significant association between disease risk scores and transcript types. In the majority of studies, having the e14a2 transcript was associated with earlier, deeper, and higher molecular response rates. Although better event-free survival was observed in patients with the e14a2 transcript in some of the studies, the majority demonstrated that transcript type did not have an impact on progression-free and overall survival. Treatment-free remission currently is a topic of much interest, and to the authors' knowledge there are limited data with conflicting results regarding the possible effects of transcript types on the outcomes of patients after discontinuation of TKIs. Because having the e14a2 transcript appears to be related to a favorable outcome, choosing second-generation TKIs for frontline therapy might be a convenient approach in patients with chronic myeloid leukemia with the e13a2 transcript. The authors believe this finding warrants further investigation. **Cancer 2018;124:3806-3818.** © 2018 American Cancer Society.

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INTRODUCTION

In 1959, Hungerford and Nowell defined a critical chromosomal abnormality in patients with chronic myeloid leukemia (CML).¹ This abnormality was the juxtaposition of the *ABL1* gene of the 9th chromosome onto the *BCR* gene of the 22nd chromosome. Today, we know that approximately 95% of patients with CML are positive for the Philadelphia chromosome (Ph⁺), and this translocation serves as a target for the tyrosine kinase inhibitors (TKIs).

The *BCR* gene contains 4 breakpoint cluster regions termed major (*M-BCR*), minor (*m-BCR*), micro (*μ-BCR*), and nano (*v-BCR*). *M-BCR* has 5 exons termed e12 to e16 (formerly b1-b5), and in 95% of patients with Ph⁺ CML, a break occurs at this point.² After the translocation of the *ABL1* gene, a chimeric messenger RNA (mRNA) transcribed from this fusion gene is translated into the p210 protein. Usually, the e13 to e14 exons of the *M-BCR* gene and the a2 to a3 exons of the *ABL1* gene are the origin of this protein whereas fusions such as e1a2 (in which the break occurs at *m-BCR* and encodes the p190 protein) and e19a2 (in which the break occurs at *μ-BCR* and encodes the p230 protein) also are possible.^{3,4} These rare forms of *BCR-ABL1* transcript types are for the most part observed in patients with Ph⁺ acute lymphoblastic leukemia. The frequency of p190 among patients with CML is reported to be approximately 1% and these patients usually are identified as being at high risk due to their inferior responses to treatment with imatinib.⁵ In patients with Ph⁺ CML, the e13a2 (“b2a2”) and e14a2 (“b3a2”) fusion genes have the highest frequency and differ from one another by 75 base pairs. To our knowledge, the clinical impact of these extra 25 amino acids in the e14a2 transcript remains unclear. However, it is known that there is a structural difference occurring in the Src homology 1 (SH1)-, SH2-, SH3-, and DNA-binding domains of the p210 protein, which can cause an alteration in kinase activity.⁶

Chimeric fusion proteins are expressed only in CML cells and presented by major histocompatibility complex (MCH) class I and II molecules on the cell surface, and antitumor responses of cytotoxic T cells are based on these human leukocyte

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antigen (HLA) proteins in patients with CML. It previously has been shown that peptide-specific CD8-positive (CD8+) or CD4+ cytotoxic T cells proliferate as a response to HLA class I or II molecules presenting proteins encoded from e13a2 and e14a2.⁷⁻¹⁰ Bocchia et al¹¹ assessed the affinity of e13a2 and e14a2 transfusion proteins to HLA class I molecules and found that 4 peptides derived from the e14a2 breakpoint had high or intermediate affinity for HLA A3, A11, and B8 molecules, but this was not true for e13a2, which did not demonstrate any affinity for these HLA class I molecules. The authors suggested that this might play a role in the impaired CD8+ cytotoxic T-cell response observed in patients with the e13a2 transcript.¹¹

These studies have provided essential information for the development of vaccines targeting these CML-specific peptides. A peptide-specific vaccine derived from e14a2 (CMLVAX100, Originator: Memorial Sloan-Kettering Cancer Center, Developer: Breakthrough Therapeutics) was used in 16 patients with CML in the chronic phase (CML-CP) who had the e14a2 transcript and who were resistant to previous treatments (imatinib in 10 cases and interferon- α [IFN- α] in 6 cases), and 5 patients receiving imatinib achieved a complete cytogenetic response (CCyR) after 6 vaccinations, 3 of whom had an undetectable transcript amount. In addition, 2 of the 6 patients receiving CMLVAX100 plus IFN- α achieved a CCyR.¹² Jain et al¹³ also assessed the efficacy of a vaccine derived from a mixture of peptides from both the e13a2 and e14a2 sequences in 10 patients with CML-CP, all with e13a2 transcripts (including 2 patients with coexpression of e13a2 and e14a2), who were in CCyR while receiving imatinib. Vaccination in combination with imatinib therapy induced improved molecular responses, a finding that then was supported by another study demonstrating promising results with the vaccine, which was derived from the e13a2 transcript.¹⁴ Recently, Vinhas et al¹⁵ used a gold nanoconjugate (a single-stranded DNA and gold nanoparticles for protection against RNase activity) specific to the e14a2 fusion point and triggered apoptosis in *BCR-ABL1*-expressing cells, which might overwhelm the imatinib resistance. Although it was shown that the addition of these vaccines to imatinib therapy may result in improved outcomes, second-generation TKIs (2GTKIs) generally are used in the management of imatinib-resistant cases of CML in daily clinical practice.

The studies demonstrating the impact of *BCR-ABL1* transcript types on the outcomes of patients with CML receiving TKIs have had conflicting results. Hematologic parameters such as platelet and leukocyte counts, spleen size, or risk scores (including the Sokal and European

Treatment and Outcome Study [EUTOS] scores but not the Euro-Hasford score) have provided opposing results by transcription type. This also makes it difficult to predict TKI response and long-term outcomes in patients with CML with regard to different transcript types. Many studies have demonstrated favorable outcomes with TKI therapy in patients with the e14a2 transcript type, but there are some that demonstrate just the opposite. In this review, we evaluated the current available literature for the differences in the short-term and long-term outcomes of patients with CML with different *BCR-ABL1* transcript types (e13a2/e14a2 or both) while receiving TKI therapy.

Methodology

We used the PubMed database and Google Scholar for a systematic literature search by using the terms “b2a2/b3a2,” “e13a2/e14a2,” or “transcript type.” We accessed 420 articles in English through December 2017. Studies unrelated to this article, reviews, case reports, letters, and duplicates were excluded. Articles reporting other rare transcript types also were excluded. Although selected abstracts presented in the American Society of Hematology meetings relevant to this topic were included, for the most part 53 full-text articles were evaluated in detail for this review.

Molecular response classification was based on *BCR-ABL1* to control gene transcript ratios, expressed on the international reporting scale (IS), in which a major molecular response (MMR) or MR³ is defined as *BCR-ABL1*^{IS} $\leq 0.1\%$, MR⁴ is defined as *BCR-ABL1*^{IS} $\leq 0.01\%$, and MR^{4.5} is defined as *BCR-ABL1*^{IS} $\leq 0.0032\%$. Early molecular response (EMR) is defined as a *BCR-ABL1*^{IS} $\leq 10\%$ at 3 or 6 months, and deep molecular response (DMR) generally is referred to as MR⁴ or MR^{4.5}; some patients may achieve responses beyond the limit of detection of the assays used, which usually is termed a complete molecular response (CMR).¹⁶

Results and Discussion

The p190 (encoded by the e1a2 transcript) and p230 (encoded by the e19a2 transcript) proteins are relatively rare in patients with CML. In what to our knowledge is the largest study to date evaluating e1a2 transcript frequency and its clinical impact among patients with CML, only 14 were found in 1292 cases (approximately 1%). Nine of these patients had CML-CP, whereas 1 had CML in accelerated phase and 4 had CML in blast crisis. These patients had inferior TKI responses compared with patients with p210, and consequently, the authors declared these cases as high-risk patients.³ In addition to

TABLE 1. Incidence of Transcript Types According to Different Studies From Different Countries

Country	Sample Size (No.)	e13a2, %	e14a2, %	Coexpression, %
Tunisia (Bennour 2013 ²)	44	36.36	63.63	0
Argentina (Sastre 2007 ¹⁸)	53	41.7	37.5	8.3
Ecuador (Paz-y-Mino 2002 ¹⁹)	40	94.6	5.4	0
Sudan (Osman 2010 ²⁰)	43	53.5	41.9	2.3
United States (Jain 2016 ²¹)	481	42	41	18
Syria (Al-Achkar 2016 ²²)	45	46.7	51.1	NR
Germany (Hanfstein 2014 ²³)	1105	41	45	14
Brazil (Pagnano 2017 ²⁴)	170	33	55	12
India (Sharma 2010 ²⁵)	87	38	54	8
India (Polampalli 2008 ²⁶)	202	32	68	0
ELN cohort (Pfirrmann 2017 ²⁷) ^a	1494	38	49	13
Italy (Castagnetti 2017 ²⁸)	559	36	52	11
India (Mir 2015 ²⁹)	200	24	68	8
Canada (Lin 2016 ³⁰)	166	36.7	50	13.3
Poland (Prejzner 2002 ³¹)	27	29	62.3	8.2
India (Deb 2014 ³²)	80	41.25	56.25	NR
Korea (Lee 2018 ³³)	120	35	60	2.5
England (Claudiani 2017 ³⁴)	64	42	58	0
United States (Verschraegen 1995 ³⁵)	362	30.2	67.9	NR
England (Khorashad 2008 ³⁶)	319	41	46	12
Serbia (Todoric-Zivanovic 2011 ³⁷)	136	25	73.5	NR
Iran (Yaghmaie 2008 ³⁸)	75	21	62	5
India (Arun 2017 ³⁹)	1260	34.3	60	NR
Thailand (Udomsakdi-Auewarakul 2000 ⁴⁰)	99	31	61	NR
Korea (Goh 2006 ⁴¹)	548	32.34	66.74	NR
Japan (Ito 2004 ⁴²)	226	30.2	67.5	NR

Abbreviations: Coexpression, coexpression of e13a2 and e14a2 transcripts; ELN, European LeukemiaNet; NR, not reported.

^aThe ELN cohort was an international cohort from Germany, Italy, Spain, and the Netherlands.

e1a2, there are only case reports reporting the e19a2 transcript in patients with CML.^{4,17}

Because p210, which is encoded by the e13a2 and e14a2 transcripts, is the most frequent *BCR-ABL1* noted in patients with CML, this review focused mainly on the impact of these transcript types on outcomes.

Relationship between transcript types and patient characteristics

The incidence rates of the *BCR-ABL1* transcript types gathered from the full-text articles are given in Table 1.^{2,18-42} Of 26 articles, 22 (85%) showed a higher incidence of the e14a2 transcript whereas only 4 articles demonstrated a higher incidence of the e13a2 transcript. The studies conducted among patients with CML from Argentina, Ecuador, and Sudan showed a higher frequency of the e13a2 transcript but had small sample sizes (53 patients, 40 patients, and 43 patients, respectively).¹⁸⁻²⁰ It is interesting to note that in the study by Paz-y-Mino et al,¹⁹ the incidence of the e13a2 transcript was 94.6%, whereas that of e14a2 was only 5.4%. Ethnicity might play a role in these results, as well as the small sample size of the study. However, a larger cohort from the United States of 481 patients also demonstrated higher incidence rates for the e13a2 transcript type.²¹

In nearly all the studies, sex did not appear to have an impact on the transcript types, including the European LeukemiaNet (ELN) cohort, which to our knowledge has the largest sample size.^{2,22-27,43-48} However, a small study consisting of 43 patients from Sudan with CML demonstrated a possible relationship between the e13a2 transcript and male sex,²⁰ which then was confirmed by other 2 large-scale studies from Italy (559 patients)²⁸ and India (200 patients)²⁹ ($P = .03$ and $P = .05$, respectively). In addition, Lin et al³⁰ demonstrated that female sex was more frequent among patients coexpressing both the e13a2 and e14a2 transcripts ($P < .05$), although no significant difference with regard to sex was shown in the e13a2 or e14a2 transcript groups.

Age did not appear to have an impact on transcript types in the majority of the studies. Bennour et al² found that patients with the e14a2 transcript type were significantly older than patients in the e13a2 group (mean age of 56 years vs 26 years; $P = .001$). Other studies did not find a correlation between age and transcript types.^{21-24,26-29,43-47}

Relationship between transcript types and hematologic parameters and risk scores at diagnosis

Structural differences in different p210 proteins (ie, e14a2 and e13a2) may result in different tyrosine kinase activities and thus, theoretically, hematologic parameters

among patients with CML with the e13a2 and e14a2 transcript types may differ.

With regard to the association between transcript types and platelet counts at the time of diagnosis, the literature search demonstrated some contradictory results. In some studies, significantly higher platelet counts were found in patients with the e14a2 transcript compared with those with the e13a2 transcript,^{2,21,44,46-48,50-52} whereas others demonstrated no difference (Table 2).^{2,21-25,27-30,43-47,49,53,54} In addition, in the study by Jain et al,²¹ significantly higher platelet levels were observed in the patients coexpressing the transcripts compared with patients with the e13a2 transcript alone. A study from Italy with 559 patients was unable to demonstrate any difference between transcript types ($P = .251$).²⁸ However, a German study with 1105 patients found that patients with the e13a2 transcript had significantly lower platelet levels compared with those with the e14a2 transcript and those coexpressing transcripts (e13a2 plus e14a2) ($P < .001$); however, after dissecting patients according to Euro-Hasford risk scores, this statistically significant difference disappeared.²³

Many studies found no association between leukocyte counts and hemoglobin levels at the time of diagnosis and the transcript types.^{2,22,24-26,28,43-45,47,49} However, in their study, the German investigators demonstrated that patients with the e13a2 transcript had significantly higher baseline median leukocyte counts compared with patients with the e14a2 transcript ($88 \times 10^9/L$ vs $65 \times 10^9/L$; $P < .001$), but no such relationship was shown for hemoglobin levels.²³ A recently published study from Brazil also reported similar findings and patients with the e13a2 transcript had a significantly higher median leukocyte count at the time of diagnosis compared with patients with the e14a2 transcript ($147 \times 10^9/L$ vs $129 \times 10^9/L$; $P = .049$) (Table 2).^{2,21-25,27-30,43-47,49}

A German study demonstrated that there was no significant difference between patients having the e13a2, e14a2, and the coexpressing e13a2 plus e14a2 transcript types in terms of spleen size at the time of diagnosis.²³ Other studies from Poland, India, and Romania with smaller sample sizes also supported this finding ($P = .941$, $P = .868$, and $P = .680$, respectively).^{25,31,47} Although having a larger spleen at the time of diagnosis was found to have a negative impact on achieving a MMR³³ and on event-free survival (EFS),²¹ to our knowledge neither study investigated the difference in spleen size among different transcript groups.

The Sokal, Euro-Hasford, EUTOS, and EUTOS Long-Term Survival (ELTS) risk scores have important

predictive value, especially for long-term outcomes. These scoring systems are based on different features, including platelet and blast counts, spleen size, and patient age. The Sokal and Euro-Hasford risk classification systems were developed before the introduction of TKIs, whereas the EUTOS and then ELTS systems were developed within the era of TKIs.^{55,56} The EUTOS score used the percentage of basophils and spleen size to divide patients into 2 groups as high risk and low risk, and 5-year progression-free survival (PFS) was found to be significantly better in the low-risk compared with the high-risk group (90% vs 82%; $P = .006$).⁵⁵ Older patient age, more peripheral blasts, a larger spleen size, and low platelet counts were found to be significantly associated with increased probabilities of dying of CML and resulted in a new long-term survival score, the so-called "ELTS score," with 3 prognostic groups. Compared with the low-risk group, patients in the intermediate-risk and high-risk groups had significantly higher probabilities of dying of CML.⁵⁶ This new score differentiated the probabilities of dying of CML better than the Sokal, Euro-Hasford, and EUTOS scores.

Lucas et al⁵⁷ found no significant difference in the distribution of Sokal risk scores between the e13a2, e14a2, and e13a2 and e14a2 coexpression groups. This also was supported by the studies by Jain et al²¹ (481 patients) and Prejzner et al³¹ (61 patients), in which there were no differences noted in terms of Sokal risk groups among patients with different transcript types ($P = .53$ and $P = .734$, respectively). Studies from India and Thailand demonstrated similar results according to the Sokal and Euro-Hasford scores^{25,54}; however, Deb et al³² demonstrated that patients with the e13a2 transcript had higher baseline Sokal and EUTOS risk scores compared with those without this transcript type ($P < .05$).

In the study by Castagnetti et al,²⁸ which included 559 patients, there were no differences noted in terms of the distribution of the Sokal, Euro-Hasford, and EUTOS risk scores among patients with the e13a2 and e14a2 transcript types ($P = .525$, $P = .322$, and $P = .662$, respectively). Similarly, in the study by Pagnano et al,²⁴ the distribution of EUTOS and Sokal risk scores was similar in patients with the e13a2, e14a2, and e13a2 plus e14a2 transcript types (Table 3).^{21,24,25,28,32,57} Pfirrmann et al²⁷ did not find any significant difference in terms of the distribution of ELTS scores between the different transcript types.

Distribution of responses to first-line imatinib according to different transcript types

In a study consisting of 481 patients with CML-CP, the distribution of the e13a2, e14a2, and e13a2 plus e14a2

TABLE 2. Distribution of Hematologic Parameters According to Transcript Types

Reference	Median/Mean Age, Years			Male Sex, %			Median Platelet Count, ×10 ⁹ /L			Median WBC, ×10 ⁹ /L			Median Hgb, g/dL								
	e13a2	e14a2	Co expression	P ^a	e13a2	e14a2	Co expression	P ^a	e13a2	e14a2	Co expression	P ^a	e13a2	e14a2	Co expression	P ^a					
Bennour 2013 ²	26.56	56.61	NR	.001	62.5	57.1	NR	NR	.49	207	681.19	NR	.001	264.7	181.5	NR	.153	9.31	10.83	NR	.078
Jain 2016 ²¹	47	49	52	.11	NR	NR	NR	NR	NR	288	405	358	.001	31	28	30	.94	12	12	13	.32
Al-Achkar 2016 ²²	39	36	NR	.66	57.1	56.5	NR	NR	1	293	257	NR	.44	143.1	115	NR	.64	11.2	10.2	NR	.28
Harfstein 2014 ²³	52	53	NR	NS	59.9	59.5	NR	NR	NS	296	430	NR	NS	87.7	65.3	NR	<.001	12.2	12.5	NR	NS
Pagnano 2017 ²⁴	43	52	48	.06	66	54	60	60	.36	343.3	431	378	.09	110	112	97.9	.7	11.7	12.1	11.6	.67
Sharma 2010 ²⁵	36.58	37.54	NR	.706	70.5	66.03	NR	NR	.196	340	360	360	.712	77	83	83	.923	11.8	12	12	.69
Kiani 2016 ⁴³	46.38	48.91	NR	NS	55.5	37.83	NR	NR	NS	364.5	392.9	NR	NS	131.8	126.8	NR	NS	11.12	10.84	NR	NS
Balatzenko 2011 ⁴⁴	49.9	51	NR	NS	65	47.45	NR	NR	NS	440.4	791.3	NR	.007	132.4	119.5	NR	NS	10.79	11.6	NR	NS
Rostami 2017 ⁴⁵	52	45	NR	.73	56	51.4	NR	NR	.93	379	386	NR	.77	140	120	NR	.07	NR	NR	NR	NR
Vasconcelos 2017 ⁴⁶	NR	NR	NR	.51	63.2	55.3	NR	NR	.28	287	429	NR	.005	147	129	NR	.049	10.5	10.5	NR	.93
Szanto 2014 ⁴⁷	51.4	50.06	NR	.9	66.6	50	NR	NR	.47	261.7	412.8	NR	.003	105.9	176	NR	.247	11.8	11.2	NR	.089
Castagnetti 2017 ²⁸	52	52	NR	.374	66	57	NR	NR	.05	293	401	NR	.251	61.6	52.2	NR	.174	12	12.3	NR	.413
Mir 2015 ²⁹	NR	NR	NR	NR	68.75	62.5	62.5	62.5	.03	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
Lin 2016 ³⁰	54	60	62.5	NS	52.4	54.2	59	59	NS	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
Mondal 2006 ⁴⁹	30.5	40	NR	NS	NR	NR	NR	NR	NR	373	250	NR	NS	162	120	NR	NS	NR	NR	NR	NR
Pfirschmann 2017 ²⁷	52	51	51	NS	59	58	62	62	NS	295	420	407	NR	NR	NR	NR	NR	NR	NR	NR	NR

Abbreviations: Coexpression, coexpression of e13a2 and e14a2 transcripts; Hgb, hemoglobin; NR, not reported; NS, not significant; WBC, white blood cell count.
^a Bold type indicates statistical significance.

TABLE 3. Sokal, Euro-Hasford, and EUTOS Risk Scores in Different Transcript Types

		e13a2,	e14a2,	Coexpression,	
Reference	Risk	No.	No.	No.	<i>P</i> ^a
Sokal					
Jain 2016 ²¹	High	11	11	9	.53
	Intermediate	47	48	21	
	Low	142	137	55	
Pagnano 2017 ²⁴	High	18	16	10	.06
	Intermediate	15	31	4	
	Low	13	25	4	
Sharma 2010 ²⁵	High	4	6	NR	.824
	Intermediate	7	14	NR	
	Low	23	33	NR	
Castagnetti 2017 ²⁸	High	43	72	NR	.525
	Intermediate	74	110	NR	
	Low	86	108	NR	
Lucas 2009 ⁵⁷	High	21	9	12	NS
	Intermediate	14	5	9	
	Low	15	6	9	
Deb 2014 ³²	High	5	7	NR	.03
	Intermediate	17	15	NR	
	Low	11	25	NR	
Euro-Hasford					
Sharma 2010 ²⁵	High	4	5	NR	.849
	Intermediate	8	10	NR	
	Low	22	38	NR	
Castagnetti 2017 ²⁸	High	12	24	NR	.322
	Intermediate	101	142	NR	
	Low	90	124	NR	
Deb 2014 ³²	High	4	1	NR	.24
	Intermediate	15	22	NR	
	Low	14	24	NR	
EUTOS					
Pagnano 2017 ²⁴	High	7	9	4	.45
	Low	47	81	16	
Castagnetti 2017 ²⁸	High	15	23	NR	.662
	Low	188	267	NR	
Deb 2014 ³²	High	17	20	NR	.027
	Low	16	27	NR	

Abbreviations: Coexpression, coexpression of e13a2 and e14a2 transcripts; EUTOS, European Treatment and Outcome Study; NR, not reported; NS, not significant.

^a Bold type indicates statistical significance.

transcript types were 200 patients (42%), 196 patients (41%), and 85 patients (18%), respectively. Patients were divided into 4 groups according to the frontline TKI therapy received as imatinib at a dose of 400 mg, imatinib at a dose of 800 mg, dasatinib 50 mg twice daily or 100 mg daily, and nilotinib 800 mg/day. Cumulative MMR and MR^{4.5} rates were significantly inferior in the e13a2 group compared with the e14a2 and coexpression groups in all treatment arms ($P = .0001$ and $P = .00001$, respectively) (Table 4).^{21,23,24,26,28-30} With regard to CCyR, patients with the e13a2 transcript who received imatinib at a dose of 400 mg/day had inferior response rates (77%) compared with other TKIs (90%-95%), but this trend was not observed in patients with the e14a2 or coexpression of the e13a2 plus e14a2 transcripts, in whom the CCyR rate

with imatinib at a dose of 400 mg/day was similar (93%) to that of other treatment modalities (93%-96%).²¹ In addition, time to achieve MMR and MR^{4.5} was longer for e13a2 cohort, but it was similar for CCyR in all transcript groups. The transcript levels declined more slowly after 3 months and 6 months of TKI therapy in the e13a2 group compared with the e14a2 and coexpression groups. When treatment responses according to different transcript types were evaluated for each different TKI treatment modality, patients with the e13a2 transcript receiving imatinib at a dose of 400 mg/day had significantly inferior CCyR, MMR, and MR^{4.5} rates compared with those observed among both patients with the e13a2 transcript who were receiving other TKI modalities and patients with other transcript types who were receiving imatinib at a dose of 400 mg/day.²¹ Lower response rates for CCyR, MMR, and MR^{4.5} in patients with the e13a2 transcript persisted, even at 60 months. The MR^{4.5} response sustainability was lower in patients with the e13a2 transcript compared with patients with the e14a2 transcript and transcript coexpression ($P = .0021$). In addition, as expected, patients treated with standard-dose imatinib (400 mg/day) were found to have the longest time to achieve MMR and MR^{4.5} irrespective of the type of transcripts, with patients with the e13a2 transcript generally having longer times to MMR and MR^{4.5} across all treatment types.²¹

In the study by Castagnetti et al,²⁸ patients receiving frontline imatinib were evaluated, and CCyR rates at 12 months for patients with the e13a2 and e14a2 transcripts were 75% and 79%, respectively ($P = .274$) (Table 5).^{21,23-25,28,57} The median time to achieve CCyR was 6 months for both groups, but patients having the e13a2 transcript type had significantly inferior MMR rates at 18 months and MR⁴ rates at 36 months compared with patients with the e14a2 transcript (52% vs 67% [$P = .001$] and 20% vs 30% [$P = .013$], respectively). In addition, the median times to achieve MMR and MR⁴ were significantly longer for patients with the e13a2 transcript compared with those with the e14a2 transcript type (12 months vs 6 months [$P = .001$] and 61 months vs 41 months [$P = .001$], respectively).²⁸

Among a patient cohort consisting of 1105 patients from Germany with CML, patients were divided into treatment arms of imatinib at a dose of 400 mg (300 patients), imatinib at a dose of 400 mg plus IFN- α (331 patients), imatinib at a dose of 400 mg plus cytarabine (150 patients), and imatinib at a dose of 800 mg (324 patients).²³ At the time of total sample analysis, a cumulative MMR incidence for the e14a2 group was better than that of patients with the e13a2 transcript ($P = .002$).

TABLE 4. Distribution of MMR, MR⁴, and MR^{4.5} Rates According to Different Transcript Types

Reference	MMR																	
	3 Months				6 Months				12 Months				Cumulative					
	e13a2, %	e14a2, %	Coexpression, %	P ^a	e13a2, %	e14a2, %	Coexpression, %	P ^a	e13a2, %	e14a2, %	Coexpression, %	P ^a	e13a2, %	e14a2, %	Coexpression, %	P ^a	Duration	
Jain 2016 ²¹	27	49	50	<.05	42	67	70	<.05	55	83	76	<.05	79	91	95	.0001	5 y	
Hantstein 2014 ²³	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	81	85	NR	.002	5 y
Pagnano 2017 ²⁴	60	84	75	.02	51	72	53	.06	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR
Polampalli 2008 ²⁶	NR	NR	NR	NR	41	48	NR	NS	76	55	NR	NS	NR	NR	NR	NR	NR	NR
Castagnetti 2017 ²⁸	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	83	88	NR	<.01	80 mo
Mir 2015 ²⁹	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	64	72.1	NR	.04	NR
Lin 2016 ³⁰	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	60.7	77.1	81.8	<.05	2 y

Reference	Cumulative MR ⁴ to MR ^{4.5b}			Duration
	e13a2, %	e14a2, %	Coexpression, %	
Jain 2016 ²¹	57	79	80	<.001
Hantstein 2014 ²³	58	76	NR	<.001
Castagnetti 2017 ²⁸	52	67	NR	.001

Abbreviations: Coexpression, coexpression of e13a2 and e14a2 transcripts; IS, international reporting scale; MR^d, BCR-ABL1^{IS}/ABL1 ≤ 0.01%; MR^{t5}, BCR-ABL1^{IS}/ABL1 ≤ 0.0032%; NR, not reported; NS, not significant.

^a Bold type indicates statistical significance.

²⁰MR^{4.5} for Jain et al²¹; MR⁴ for others.

TABLE 5. CCyR Rates in Different Transcript Types Shown in the Literature

Reference	CCyR																Duration		
	3 Months				6 Months				12 Months				Cumulative						
	e13a2, %	e14a2, %	Coexpression, %	<i>P</i> ^a	e13a2, %	e14a2, %	Coexpression, %	<i>P</i> ^a	e13a2, %	e14a2, %	Coexpression, %	<i>P</i> ^a	e13a2, %	e14a2, %	Coexpression, %	<i>P</i> ^a			
Jain 2016 ²¹	59	67	63	NR	NR	73	81	82	NR	NR	NR	NR	NR	89	94	94	NS	60 mo	
Hanfstein 2014 ²³	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	94.6	93.3	93.6	NS	5 y
Pagnano 2017 ²⁴	NR	NR	NR	NR	NR	43	70	64	.02	62	78	79	.16	NR	NR	NR	NR	NR	NR
Sharma 2010 ²⁵	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	61	35	NR	.396	2 y	NR
Castagnetti 2017 ²⁸	NR	NR	NR	NR	NR	NR	NR	NR	NR	75	79	NR	.274	89	88	NR	.916	80 mo	NR
Lucas 2009 ⁵⁷	NR	NR	NR	NR	NR	NR	NR	NR	NR	25	53.8	NR	.01	NR	NR	NR	NR	NR	NR

Abbreviations: CCyR, complete cytogenetic response; Coexpression, coexpression of e13a2 and e14a2 transcripts; NR, not reported; NS, not significant.

^a Bold type indicates statistical significance.

DMR (MR⁴ or deeper) rates also were found to be superior in patients with the e14a2 transcript compared with patients with the e13a2 transcript (76% vs 58%; $P < .001$). Patients coexpressing both transcript types demonstrated no difference in MMR rates when compared with patients with the e13a2 or e14a2 transcripts, but this group had significantly lower MR⁴ rates compared with patients with the e14a2 transcript ($P = .004$) (Table 4).^{21,23,24,26,28-30} When evaluating the treatment arms separately, in the patients treated with imatinib at a dose of 400 mg plus IFN- α , those patients with the e14a2 transcript type achieved significantly higher MMR rates when compared with the e13a2 group ($P = .004$). In addition, MR⁴ rates also were superior among patients with the e14a2 transcript compared with those with the e13a2 transcript among the patients treated with imatinib at a dose of 400 mg plus IFN- α , imatinib at a dose of 400 mg plus cytarabine, and imatinib at a dose of 800 mg ($P < .001$, $P = .004$, and $P = .028$, respectively). However, there were no significant differences in the cumulative incidence of major cytogenetic response, CCyR, PFS, and overall survival (OS) noted between different transcript types in all treatment arms, and the authors concluded that the transcript types did not serve as a useful prognostic tool with which to predict long-term outcomes, at least in this cohort of patients.²³

A retrospective study from Canada with 166 patients with CML-CP demonstrated that MMR rates in the e14a2 and coexpression groups were both significantly higher than that of patients with the e13a2 transcript (77.1% and 81.8% vs 60.7%, respectively; P values not given) (Table 4).^{21,23,24,26,28-30} The median time to achieve MMR did not differ between the groups, and similarly the disease progression rates and median time to disease progression were comparable. The authors concluded that having an e14a2 transcript type was associated with better molecular responses.³⁰ Another study from India with 200 patients supported the findings of the Canadian study²⁹ by demonstrating that the MMR rates of patients with the e14a2, e13a2, and coexpression transcripts were 72.1%, 24.7%, and 3%, respectively ($P = .04$). Complete hematologic response and MMR rates also were similar to what was observed in the study by Lin et al,³⁰ and also were superior in patients with the e14a2 transcript compared with those with the e13a2 transcript ($P = .05$ for complete hematologic response and $P = .04$ for MMR).²⁹ However, in this study, approximately 42% of patients were in accelerated phase or blast crisis, which was not the case in the Canadian study.²⁹

A retrospective study evaluated 170 treatment-naïve patients who were treated with imatinib at a dose of 400 or 800 mg/day as frontline therapy and compared responses according to transcript type within the entire cohort regardless of the daily imatinib dose. CCyR rates at 6 months were 43% for the e13a2 group, 70% for patients with the e14a2 transcript, and 64% for those demonstrating coexpression ($P = .02$). However, no such difference was observed at 12 months of TKI therapy ($P = .16$). EMR assessment at 3 months also favored patients with the e14a2 transcript ($P = .02$).²⁴ In the study by Lucas et al,⁵⁷ patients having the e14a2 transcript type had superior CCyR rates and achieved this response faster when compared with the e13a2 group ($P = .01$ and $P = .006$, respectively) in a cohort of 78 patients with CML receiving first-line imatinib (Table 5).^{21,23-25,28,57}

Lee et al³³ divided a total of 120 patients who did not achieve EMR at 3 months into 3 groups as patients with the e13a2 transcript (group 1), those with the e14a2 transcript with a spleen measuring >9 cm (group 2), and those with the e14a2 transcript with a spleen measuring ≤ 9 cm (group 3). While receiving frontline imatinib treatment, overall MMR rates in these 3 patient groups were 19%, 20.8%, and 56.1%, respectively, and the authors concluded that having the e13a2 transcript type and splenomegaly >9 cm were negative predictors of achieving MMR.³³

The study by Vega-Ruiz et al⁵⁸ reported similar results, in which the authors evaluated imatinib response in 251 newly diagnosed patients and 229 patients after IFN- α failure. Among the treatment-naïve group, MMR and CMR rates (defined as undetectable transcript levels) were superior in patients with the e14a2 transcript compared with those with the e13a2 transcript (59% vs 77% [$P = .008$] and 25% vs 47% [$P = .002$], respectively). Improved MMR and CMR rates also were observed in the IFN- α failure group for patients having the e14a2 transcript compared with patients with the e13a2 transcript (34% vs 63% [$P = 0.001$] and 16% vs 42% [$P = 0.001$], respectively).⁵⁸

Similarly, among patients with CML-CP receiving imatinib both in the upfront setting or after first-line IFN- α , Bonifacio et al⁵⁹ showed that having the e14a2 transcript was associated with durable MR⁴ when compared with those with the e13a2 transcript ($P = .003$).⁵⁹

Although in the majority of the publications it was stated that the patients with the e14a2 transcript usually achieved better responses and had favorable outcomes, there also are some studies with contradictory results. A study from India with 87 patients with CML

demonstrated that those with the e13a2 transcript had better responses while receiving imatinib compared with patients having the e14a2 transcript. CCyR rates were 59% versus 28%, respectively, in favor of the e13a2 transcript type, and the difference was statistically significant ($P = .04$). However among this cohort, there were patients with a prior treatment other than imatinib (hydroxyurea or IFN- α), and there were only 30 treatment-naïve patients. After performing the analysis within these treatment-naïve patients, the CCyR rates among the different transcript groups were similar ($P = .396$).²⁵ Polampalli et al²⁶ found no difference in terms of MMR and CCyR rates at 6 months and 12 months between patients with the e13a2 and e14a2 transcript types, but there were more patients with the e13a2 transcript who progressed to myeloid blast crisis (Table 4).^{21,23,24,26,28-30}

Distribution of responses under 2GTKIs according to different transcript types in the frontline setting

To the best of our knowledge, there are limited data evaluating the responses and outcomes under first-line 2GTKIs according to *BCR-ABL1* transcript types.

In a study from The University of Texas MD Anderson Cancer Center (MDACC), there were 105 patients receiving upfront dasatinib 50 mg twice daily or 100 mg daily and 108 patients receiving frontline nilotinib 400 mg twice daily.²¹ In patients with the e13a2 transcript, overall CCyR rates for both 2GTKIs were superior to that of imatinib at a dose of 400 mg/day but comparable to that of imatinib at a dose of 800 mg/day (77% for imatinib at a dose of 400 mg/day vs 90%-95% for other treatment arms; P value not given). Overall MMR and MR^{4.5} rates among patients with the e13a2 transcript receiving treatment with 2GTKIs also were higher than those for patients receiving imatinib at doses of 400 mg/day and 800 mg/day (P value not given). For MR³ and MR^{4.5}, patients with the e13a2 transcript who were treated with imatinib at a dose of 400 mg/day had a trend toward an inferior response rate compared with those treated with other TKI modalities. In addition, MR³ and MR^{4.5} rates were comparable between all TKI groups for patients with the e14a2 transcript, with the exception of patients treated with nilotinib 800 mg daily, who demonstrated an inferior MR^{4.5} rate in both the e13a2 and e14a2 cohorts compared with patients treated with imatinib at a dose of 800 mg/day or dasatinib 2 \times 50 mg/day or 100 mg daily.²¹ The MR^{4.5} rate of nilotinib 400 mg twice daily in those patients with the e14a2 transcript was found to be inferior to that of imatinib at a dose of 800 mg/day and dasatinib 50 mg twice daily or 100 mg daily (64% for imatinib at a

dose of 400 mg/day, 85% for imatinib at a dose of 800 mg/day, 89% for dasatinib 50 mg twice daily or 100 mg/day, and 68% for nilotinib 800 mg/day). The CCyR and MMR rates of patients with the e14a2 transcript receiving first-line nilotinib 400 mg twice daily treatment were comparable to those of other treatment arms. Based on these results, the authors proposed a possible frontline therapy approach of 2GTKIs for patients with the e13a2 transcript and imatinib at a dose of 400 mg/day for patients with the e14a2 transcript.²¹ They also stated that having the e14a2 transcript type (compared with patients with the e13a2 transcript, but not the coexpressing patients), receiving first-line treatment with imatinib at a dose of 800 mg/day or 2GTKIs, and having a spleen measurement of <10 cm at the time of the initial presentation have prognostic value for EFS. Positive predictors for treatment-free remission (TFR) were defined as having the e14a2 transcript or coexpressing the e13a2 plus e14a2 transcripts, and frontline treatments with imatinib at a dose of 800 mg daily or dasatinib 50 mg twice daily or 100 mg/day. However, the only positive predictor found for OS was having both the e13a2 and e14a2 transcripts.²¹ The authors also evaluated patients according to ELN optimal response criteria and found that having the e14a2 transcript or coexpressing the e13a2 plus e14a2 transcripts were positive predictors for achieving MMR at 6 months and 12 months of TKI treatment. In addition, the e14a2 transcript had a prognostic value for superior major cytogenetic response at 3 months and improved CCyR at 6 months of TKI treatment.²¹

Castagnetti et al⁴⁸ demonstrated that the cumulative MMR (82% vs 88%; $P = .135$), MR⁴ (60% vs 69%; $P = .101$), estimated PFS (88% vs 93%; $P = .547$), and estimated OS (89% vs 94%; $P = .436$) rates were comparable between patients having the e13a2 and e14a2 transcripts, respectively, among 328 patients with CML-CP receiving upfront nilotinib. Although there was a trend toward better responses and outcomes in patients with the e14a2 transcript, none reached statistical significance.⁴⁸

In another study from MDACC, the authors evaluated the distribution of responses and outcomes among 85 patients (47 with recurrent/refractory disease and 38 newly diagnosed individuals) receiving ponatinib (a third-generation TKI) according to transcript type. In the patients with recurrent/refractory disease, the overall CCyR and MMR rates of patients with the e13a2, e14a2, and coexpression transcripts were 50%, 61%, and 50%, respectively, and 29%, 52%, and 30%, respectively.⁶⁰ While receiving frontline ponatinib, the median *BCR-ABL1*^{IS} levels at 3 months were at least MR³ for all

TABLE 6. Long-Term Outcomes and Survival Data According to Transcript Types

Reference	e13a2, %	e14a2, %	Coexpression, %	<i>P</i> ^a	Duration
OS					
Jain 2016 ²¹	88	95	98	.34	5 y
Pagnano 2017 ²⁴	96	88	NR	NS	5 y
Pagnano 2017 ²⁴	94	76	67	.03	10 y
Pfirschmann 2017 ²⁷	89	93	93	.106	5 y
Castagnetti 2017 ²⁸	83	90	NR	.017	7 y
EFS					
Jain 2016 ²¹	79	89	87	.09	5 y
Pagnano 2017 ²⁴	82	71	71	.41	10 y
PFS					
Pagnano 2017 ²⁴	94	89	75	.13	10 y
Castagnetti 2017 ²⁸	81	89	NR	.005	7 y
FFS					
Castagnetti 2017 ²⁸	54	71	NR	<.001	7 y
TFS					
Jain 2016 ²¹	91	97	99	0.01	5 y

Abbreviations: Coexpression, coexpression of e13a2 and e14a2 transcripts; EFS, event-free survival; FFS, failure-free survival; NR, not reported; NS, not significant; OS, overall survival; PFS, progression-free survival; TFS, transformation-free survival.

^a Bold type indicates statistical significance.

transcript groups, and at 6 months all groups showed an MR⁴ or deeper response with no significant differences noted between different transcript groups. The 3-year probability of failure-free survival (FFS) was comparable for all groups, but patients with the e13a2 transcript demonstrated inferior results with regard to the 3-year probability of OS when compared with patients with the e14a2 and e13a2 plus e14a2 transcripts ($P = .08$ and $P = .03$, respectively).⁶⁰

Long-term outcomes and survival according to different transcript types

Long-term outcomes and survival data of the studies are summarized in Table 6.^{21,24,27,28} In a recently published study in which 1494 patients who were treated with first-line imatinib-based regimens were included, there was no significant difference in 5-year OS noted between patients with the e13a2, e14a2, and coexpressing transcripts when patients were stratified according to ELTS risk scores (89%, 93%, and 93%, respectively; $P = .106$). In addition, the probability of dying of CML-related causes was similar in patients with the e13a2, e14a2, and e13a2 plus e14a2 transcripts (5%, 3%, and 2%, respectively; $P = .256$).²⁷

In contrast to the previous study, Castagnetti et al²⁸ demonstrated that in patients with the e13a2 and e14a2 transcript types, the 7-year OS rates while receiving imatinib at a dose of 400 mg/day were 83% versus 91%,

whereas in the high-dose (800 mg/day) imatinib group, the 7-year OS rates were 82% versus 87% ($P = .038$ and $P = .232$, respectively), demonstrating that patients with the e14a2 transcript had a significantly higher 7-year OS compared with patients with the e13a2 transcript receiving standard-dose imatinib. In addition, patients with the e14a2 transcript had significantly better PFS for both daily imatinib doses ($P = .027$ for imatinib at a dose of 400 mg/day and $P = .039$ for imatinib at a dose of 800 mg/day), and the FFS rates were significantly superior in patients with the e14a2 transcript type for both the standard-dose and high-dose groups ($P = .004$ for imatinib at a dose of 400 mg/day and $P = .011$ for imatinib at a dose of 800 mg/day).²⁸ That said, in the same study, the authors suggested that transcript type may be able to predict survival regardless of the daily imatinib dose administered and that patients with the e14a2 transcript had significantly superior OS, PFS, and FFS when compared with patients with the e13a2 transcript ($P = .017$, $P = .005$, and $P < .001$, respectively).²⁸

There were no differences in terms of 5-year EFS and OS noted between patients with the e13a2, e14a2, and coexpressing transcripts in the MDACC cohort.²¹ However, patients with the e14a2 or coexpressed e13a2 plus e14a2 transcripts and who achieved optimal responses at 3 months of TKI therapy according to the 2013 ELN response criteria had better transformation-free survival rates than patients with the e13a2 transcript (95%, 99%, and 89%, respectively; $P = .033$).

As stated earlier, in the Korean study,³³ the cohort was divided into 3 groups, and patients in group 3 were found to have significantly higher MMR rates and better 5-year FFS compared with the other groups, but there was no significant difference with regard to OS and PFS noted between all groups. Another study with a relatively shorter follow-up period also demonstrated similar results, with no significant difference noted in the 2-year OS despite a better CCyR in patients with the e14a2 transcript.⁵⁷

In the study by Pagnano et al,²⁴ there were no significant differences noted in terms of 5-year EFS, PFS, and OS between the transcript groups. Despite significantly better EMR ($BCR-ABL1^{IS} \leq 10\%$ at 3 months) and CCyR rates being observed in patients with the e14a2 transcript, the 10-year OS in patients with the e13a2 transcript was superior to that of patients with the e14a2 transcript and the coexpression group ($P = .03$).²⁴ Although there was no significant difference between the patient groups with regard to median age, patients in the e13a2 cohort were younger, and the authors concluded that the favorable OS detected in these individuals most likely was due to this age difference.²⁴

Impact of *BCR-ABL1* transcript types on TKI discontinuation and TFR

Discontinuation of TKI therapy and TFR currently are topics of much interest for physicians who treat patients with CML, and to our knowledge the impact of transcript types on the outcome of TFR is not yet fully understood. Lee et al⁶¹ assessed the effect of transcript types on sustained MMR rates and CMR at 12 months of imatinib cessation in the Korean Imatinib Discontinuation Study (KIDS). They demonstrated that sustained MMR rates and CMR at 12 months were comparable for both patients with the e13a2 and e14a2 transcript types ($P = .977$ and $P = .859$, respectively).⁶¹

In a recent study, Claudiani et al³⁴ analyzed 37 patients with the e14a2 transcript and 27 patients with the e13a2 transcript who achieved and maintained MR⁴ or MR^{4.5} for at least 12 months and then discontinued TKI therapy. Thirty-two patients received imatinib, 17 patients were treated with nilotinib, and the remaining 15 patients were receiving dasatinib at the time of TKI cessation. Thirteen patients received upfront 2G TKI treatment. After TKI discontinuation, 37 patients (58%) sustained molecular remission at a median of 26 months, and patients with the e13a2 transcript demonstrated inferior results compared with the e14a2 group (45% vs 70%).³⁴ The 3-year probability of TFR was 53% for the entire cohort, and this percentage was higher for patients with the e14a2 transcript compared with those with the e13a2 transcript (66% vs 38%, respectively). Having the e14a2 transcript and being aged ≥ 40 years at the time of diagnosis were marked as positive predictors of TFR ($P = .016$ for the e14a2 vs e13a2 transcript and $P = .003$ for age ≥ 40 years vs age < 40 years).³⁴

Conclusions

TKI therapy has revolutionized the management of patients with CML-CP, but some patients still have inferior responses and worse long-term outcomes. There are many factors that might play a role, including the different *BCR-ABL1* transcript types at baseline. In this review, we evaluated the current literature regarding the impact of different transcripts (e13a2, e14a2, or coexpression of e13a2 plus e14a2) on the short-term and long-term outcomes as well as the correlation of these transcript types with the disease characteristics at the time of the initial diagnosis. In approximately one-half of the studies, the e14a2 transcript was associated with higher platelet counts, whereas other studies did not demonstrate such an association. Almost no studies demonstrated a significant association between disease risk score and *BCR-ABL1*

transcript type. In the majority of the studies, having the e14a2 transcript at baseline was associated with higher molecular response rates (including EMR and DMR), whereas some studies demonstrated just the opposite. For the long-term outcomes, although some of the studies demonstrated better EFS in patients with the e14a2 transcript, the majority of studies demonstrated that transcript type does not have an impact on PFS and OS. TFR is a novel topic for discussion in the management of patients with CML, and to our knowledge there are limited data with conflicting results regarding the possible effects of transcript type on the outcomes of patients after TKI discontinuation.

Because having the e14a2 transcript appears to be related to favorable outcomes, choosing alternative therapies such as 2GTKIs in the frontline setting might be a convenient approach in patients with the e13a2 transcript, which generally is associated with an inferior outcome, and we believe this warrants further investigation. Prospective and randomized controlled trials with larger sample sizes still are needed to determine the impact of transcript type on the short-term and long-term outcomes in patients with CML who are receiving TKI therapy.

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A. Emre Eskazan has received honoraria for advisory board membership from Novartis for work performed outside of the current study, and has received lecture fees from Novartis and Bristol-Myers Squibb for work performed outside of the current study.

REFERENCES

- Nowell C. The minute chromosome (Ph 1) in chronic granulocytic leukemia. *Ann Hematol*. 1962;8:65-66.
- Bennour A, Ouahchi I, Achour B, et al. Analysis of the clinico-hematological relevance of the breakpoint location within M-BCR in chronic myeloid leukemia. *Med Oncol*. 2013;30:348.
- Duan MH, Li H, Cai H. A rare e13a3 (b2a3) BCR-ABL1 fusion transcript with normal karyotype in chronic myeloid leukemia: the challenges in diagnosis and monitoring minimal residual disease (MRD). *Leuk Res*. 2017;59:8-11.
- Sazawal S, Chikkara S, Singh K, et al. Chronic myeloid leukemia with a rare fusion transcript, e19a2 BCR-ABL1: a report of three cases from India. *Ann Diagn Pathol*. 2017;27:24-27.
- Verma D, Kantarjian HM, Jones D, et al. Chronic myeloid leukemia (CML) with P190BCR-ABL: analysis of characteristics, outcomes, and prognostic significance. *Blood*. 2009;114:2232-2235.
- Hai A, Kizilbash NA, Zaidi SH, Alruwaili J, Shahzad K. Differences in structural elements of Bcr-Abl oncoprotein isoforms in chronic myelogenous leukemia. *Bioinformation*. 2014;10:108-114.
- Pinilla-Ibarz J, Korontsvit T, Zakhaleva V, Roberts W, Scheinberg DA. Synthetic peptide analogs derived from bcr/abl fusion proteins and the induction of heteroclitic human T-cell responses. *Haematologica*. 2005;90:1324-1332.
- Mannering S, McKenzie J, Fearnley D, Hart DN. HLA-DR1-restricted bcr-abl (b3a2)-specific CD4+ T lymphocytes respond to dendritic cells pulsed with b3a2 peptide and antigen-presenting cells exposed to b3a2 containing cell lysates. *Blood*. 1997;90:290-297.
- Yasukawa M, Ohminami H, Kaneko S, et al. CD4(+) cytotoxic T-cell clones specific for bcr-abl b3a2 fusion peptide augment colony formation by chronic myelogenous leukemia cells in a b3a2-specific and HLA-DR-restricted manner. *Blood*. 1998;92:3355-3361.
- Bosch GJ, Joosten AM, Kessler JH, Melief CJ, Leeksa OC. Recognition of BCR-ABL positive leukemic blasts by human CD4+ T cells elicited by primary in vitro immunization with a BCR-ABL breakpoint peptide. *Blood*. 1996;88:3522-3527.
- Bocchia M, Wentworth P, Southwood S, et al. Specific binding of leukemia oncogene fusion protein peptides to HLA class I molecules. *Blood*. 1995;85:2680-2684.
- Bocchia M, Gentili S, Abruzzese E, et al. Effect of a p210 multipptide vaccine associated with imatinib or interferon in patients with chronic myeloid leukaemia and persistent residual disease: a multicentre observational trial. *Lancet*. 2005;365:657-662.
- Jain N, Reuben JM, Kantarjian H, et al. Synthetic tumor-specific breakpoint peptide vaccine in patients with chronic myeloid leukemia and minimal residual disease: a phase 2 trial. *Cancer*. 2009;115:3924-3934.
- Maslak P, Dao T, Gomez M, et al. A pilot vaccination trial of synthetic analog peptides derived from the BCR-ABL breakpoints in CML patients with minimal disease. *Leukemia*. 2008;22:1613-1616.
- Vinhas R, Fernandes AR, Baptista PV. Gold nanoparticles for BCR-ABL1 gene silencing: improving tyrosine kinase inhibitor efficacy in chronic myeloid leukemia. *Mol Ther Nucleic Acids*. 2017;7:408-416.
- Dulucq S, Mahon FX. Deep molecular responses for treatment-free remission in chronic myeloid leukemia. *Cancer Med*. 2016;5:2398-2411.
- Andrikovics H, Nahajevszky S, Szilvasi A, et al. First and second line imatinib treatment in chronic myelogenous leukemia patients expressing rare e1a2 or e19a2 BCR-ABL transcripts. *Hematol Oncol*. 2007;25:143-147.
- Sastre DA, Argarana CE, Heller VB, et al. An analysis of multiplex-PCR in the detection of BCR-ABL transcripts in hematological disorders. *Genet Mol Biol*. 2007;30:520-523.
- Paz-y-Mino C, Burgos R, Morillo SA, Santos JC, Fiallo BF, Leone PE. BCR-ABL rearrangement frequencies in chronic myeloid leukemia and acute lymphoblastic leukemia in Ecuador, South America. *Cancer Genet Cytogenet*. 2002;132:65-67.
- Osman EAI, Hamad K, Elmula IM, Ibrahim ME. Frequencies of BCR-ABL1 fusion transcripts among Sudanese chronic myeloid leukaemia patients. *Genet Mol Biol*. 2010;33:229-231.
- Jain P, Kantarjian H, Patel KP, et al. Impact of BCR-ABL transcript type on outcome in patients with chronic-phase CML treated with tyrosine kinase inhibitors. *Blood*. 2016;127:1269-1275.
- Al-Achkar W, Moassass F, Youssef N, Wafa A. Correlation of p210 BCR-ABL transcript variants with clinical, parameters and disease outcome in 45 chronic myeloid leukemia patients. *J BUON*. 2016; 21:444-449.
- Hanfstein B, Lauseker M, Hehlmann R, et al; SAKK and the German CML Study Group. Distinct characteristics of e13a2 versus e14a2 BCR-ABL1 driven chronic myeloid leukemia under first-line therapy with imatinib. *Haematologica*. 2014;99:1441-1447.
- Pagnano KBB, Miranda EC, Delamain MT, et al. Influence of BCR-ABL transcript type on outcome in patients with chronic-phase chronic myeloid leukemia treated with imatinib. *Clin Lymphoma Myeloma Leuk*. 2017;17:728-733.
- Sharma P, Kumar L, Mohanty S, Kochupillai V. Response to imatinib mesylate in chronic myeloid leukemia patients with variant BCR-ABL fusion transcripts. *Ann Hematol*. 2010;89:241-247.
- Polampalli S, Choughule A, Negi N, et al. Analysis and comparison of clinicohematological parameters and molecular and cytogenetic response of two Bcr/Abl fusion transcripts. *Genet Mol Res*. 2008;7: 1138-1149.
- Pfirschmann M, Evtimova D, Sausse S, et al. No influence of BCR-ABL1 transcript types e13a2 and e14a2 on long-term survival: results in 1494 patients with chronic myeloid leukemia treated with imatinib. *J Cancer Res Clin Oncol*. 2017;143:843-850.
- Castagnetti F, Gugliotta G, Breccia M, et al; GIMEMA CML Working Party. The BCR-ABL1 transcript type influences response

- and outcome in Philadelphia chromosome-positive chronic myeloid leukemia patients treated frontline with imatinib. *Am J Hematol*. 2017;92:797-805.
29. Mir R, Ahmad I, Javid J, et al. Simple multiplex RT-PCR for identifying common fusion BCR-ABL transcript types and evaluation of molecular response of the a2b2 and a2b3 transcripts to imatinib resistance in north Indian chronic myeloid leukemia patients. *Indian J Cancer*. 2015;52:314-318.
 30. Lin HX, Sjaarda J, Dyck J, et al. Gender and BCR-ABL transcript type are correlated with molecular response to imatinib treatment in patients with chronic myeloid leukemia. *Eur J Haematol*. 2016;96:360-366.
 31. Prejzner W. Relationship of the BCR gene breakpoint and the type of BCR/ABL transcript to clinical course, prognostic indexes and survival in patients with chronic myeloid leukemia. *Med Sci Monit*. 2002;8:BR193-BR197.
 32. Deb P, Chakrabarti P, Chakrabarty S, et al. Incidence of BCR-ABL transcript variants in patients with chronic myeloid leukemia: their correlation with presenting features, risk scores and response to treatment with imatinib mesylate. *Indian J Med Paediatr Oncol*. 2014;35:26-30.
 33. Lee SE, Choi SY, Kim SH, et al. Baseline BCR-ABL1 transcript type of e13a2 and large spleen size are predictors of poor long-term outcomes in chronic phase chronic myeloid leukemia patients who failed to achieve an early molecular response after 3 months of imatinib therapy. *Leuk Lymphoma*. 2018;59:105-113.
 34. Claudiani S, Apperley JF, Gale RP, et al. E14a2 BCR-ABL1 transcript is associated with a higher rate of treatment-free remission in persons with chronic myeloid leukemia after stopping tyrosine kinase-inhibitor therapy. *Haematologica*. 2017;102:e297-e299.
 35. Verschraegen CF, Kantarjian HM, Hirsch-Ginsberg C, et al. The breakpoint cluster region site in patients with Philadelphia chromosome-positive chronic myelogenous leukemia. Clinical, laboratory, and prognostic correlations. *Cancer*. 1995;76:992-997.
 36. Khorashad JS, de Lavallade H, Apperley JF, et al. Finding of kinase domain mutations in patients with chronic phase chronic myeloid leukemia responding to imatinib may identify those at high risk of disease progression. *J Clin Oncol*. 2008;26:4806-4813.
 37. Todoric-Zivanovic B, Strnad M, Stamatovic D, et al. Frequency of BCR-ABL fusion transcripts in Serbian patients with chronic myeloid leukemia. *J BUON*. 2011;16:104-107.
 38. Yaghmaie M, Ghaffari SH, Ghavamzadeh A, et al. Frequency of BCR-ABL fusion transcripts in Iranian patients with chronic myeloid leukemia. *Arch Iran Med*. 2008;11:247-251.
 39. Arun AK, Senthamizhselvi A, Mani S, et al. Frequency of rare BCR-ABL1 fusion transcripts in chronic myeloid leukemia patients. *Int J Lab Hematol*. 2017;39:235-242.
 40. Udomsakdi-Auewarakul C, U-Pratya Y, Boonmoh S, Vatanavicharn S. Detection of molecular variants of BCR-ABL gene in bone marrow and blood of patients with chronic myeloid leukemia by reverse-transcriptase polymerase chain reaction (RT-PCR). *J Med Assoc Thai*. 2000;83:928-935.
 41. Goh HG, Hwang JY, Kim SH, Lee YH, Kim YL, Kim DW. Comprehensive analysis of BCR-ABL transcript types in Korean CML patients using a newly developed multiplex RT-PCR. *Transl Res*. 2006;148:249-256.
 42. Ito T, Tanaka H, Tanaka K, et al. Insertion of a genomic fragment of chromosome 19 between BCR intron 19 and ABL intron 1a in a chronic myeloid leukaemia patient with μ -BCR-ABL (e19a2) transcript. *Br J Haematol*. 2004;126:752-753.
 43. Kiani AA, Shahsavari F, Gorji M, Ahmadi K, Nazarabad VH, Bahmani B. Prevalence of Abelson murine leukemia viral oncogene homolog-breakpoint cluster region fusions and correlation with peripheral blood parameters in chronic myelogenous leukemia patients in Lorestan Province, Iran. *Int J Appl Basic Med Res*. 2016;6:271-275.
 44. Balatzenko G, Vundinti BR, Margarita G. Correlation between the type of bcr-abl transcripts and blood cell counts in chronic myeloid leukemia—a possible influence of *mdr1* gene expression. *Hematol Rep*. 2011;3:e3.
 45. Rostami G, Hamid M, Jalaeikhoo H. Impact of the BCR-ABL1 fusion transcripts on different responses to imatinib and disease recurrence in Iranian patients with chronic myeloid leukemia. *Gene*. 2017;627:202-206.
 46. Vasconcelos A, Azevedo I, Melo F, Neves WB, Azevedo ACAC, Melo RAM. BCR-ABL1 transcript types showed distinct laboratory characteristics in patients with chronic myeloid leukemia. *Genet Mol Res*. 2017;16(2). doi: 10.4238/gmr16029541.
 47. Szanto A, Pap Z, Denes L, et al. Analysis of the relationship between clinical parameters and the 2 variants of M-BCR-ABL fusion gene in chronic myeloid leukemia. *Romanian Journal of Functional & Clinical, Macro- & Microscopical Anatomy & of Anthropology*. 2014;13(3).
 48. Castagnetti F, Gugliotta G, Breccia M, et al. Prognostic value of BCR-ABL1 transcript type in chronic myeloid leukemia patients treated frontline with nilotinib [abstract]. *Blood*. 2016;128:3070.
 49. Mondal BC, Bandyopadhyay A, Majumdar S, et al. Molecular profiling of chronic myeloid leukemia in eastern India. *Am J Hematol*. 2006;81:845-849.
 50. Inokuchi K, Inoue T, Tojo A, et al. A possible correlation between the type of bcr-abl hybrid messenger RNA and platelet count in Philadelphia-positive chronic myelogenous leukemia [see comments]. *Blood*. 1991;78:3125-3127.
 51. Perego RA, Costantini M, Cornacchini G, et al. The possible influences of B2A2 and B3A2 BCR/ABL protein structure on thrombopoiesis in chronic myeloid leukaemia. *Eur J Cancer*. 2000;36:1395-1401.
 52. Blickstein D, Aviram A, Luboshitz J, et al. BCR-ABL transcripts in bone marrow aspirates of Philadelphia-negative essential thrombocythemia patients: clinical presentation. *Blood*. 1997;90:2768-2771.
 53. Martinez-Mancilla M, Gutierrez M, de la Rosa GZ, Garcia-Carranca A, Gariglio P, Miranda E. Younger age and shorter chronic phase in b2a2-positive chronic myeloid leukemia adults with high white blood cell count at diagnosis. *Haematologica*. 2002;87:666-668.
 54. Auewarakul CU, Huang S, Yimyam M, Boonmoh S. Natural history of Southeast Asian chronic myeloid leukemia patients with different BCR-ABL gene variants. *Acta Haematol*. 2006;116:114-119.
 55. Hasford J, Baccarani M, Hoffmann V, et al. Predicting complete cytogenetic response and subsequent progression-free survival in 2060 patients with CML on imatinib treatment: the EUTOS score. *Blood*. 2011;118:686-692.
 56. Pfirrmann M, Baccarani M, Saussele S, et al. Prognosis of long-term survival considering disease-specific death in patients with chronic myeloid leukemia. *Leukemia*. 2016;30:48-56.
 57. Lucas CM, Harris RJ, Giannoudis A, et al. Chronic myeloid leukemia patients with the e13a2 BCR-ABL fusion transcript have inferior responses to imatinib compared to patients with the e14a2 transcript. *Haematologica*. 2009;94:1362-1367.
 58. Vega-Ruiz A, Kantarjian H, Shan J, et al. Better molecular response to imatinib for patients (pts) with chronic myeloid leukemia (CML) in chronic phase (CP) carrying the b3a2 transcript compared to b2a2 [abstract]. *Blood*. 2007;110:1939.
 59. Bonifacio M, Scaffidi L, Binotto G, et al. Predictive factors of stable deep molecular response in chronic myeloid leukemia patients treated with imatinib standard dose: a study from the Gruppo Triveneto LMC [abstract]. *Blood*. 2015;126:597.
 60. Kantarjian HM, Romo CG, Luthra R, et al. BCR-ABL fusion transcripts in patients with chronic myeloid leukemia in chronic phase (CML-CP): experience with ponatinib from MD Anderson Cancer Center [abstract]. *Blood*. 2013;122:2727.
 61. Lee SE, Choi SY, Bang JH, et al. Predictive factors for successful imatinib cessation in chronic myeloid leukemia patients treated with imatinib. *Am J Hematol*. 2013;88:449-454.